## High levels of diploid male production in a primitively eusocial bee (Hymenoptera: Halictidae)

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Under single locus complementary sex determination (sl-CSD), diploid males are produced from fertilized eggs that are homozygous at the sex-determining locus. Diploid males are effectively sterile, and thus their production generates a costly genetic load. Using allozyme electrophoresis, a large number of diploid males were detected in natural populations of the primitively eusocial bee, *Halictus poeyi* Lepeletier collected in southern and central Florida during May 2000. Estimates for the proportion of diploids that are male ranged from 9.1% to 50%, while the frequency of matched matings ranged from 18.2% to 100%. The effective number of alleles at the sex-determining locus ranged from two to 11, with an average of five alleles. The effective population size of *Halictus poeyi* was estimated to be 19.6  $\pm$  2.5 SE. These data are interpreted in the light of the biogeographic history of Florida and the social biology/population dynamics of *H. poeyi*.

**Keywords:** diploid males production, effective population size, *Halictus poeyi*, Hymenoptera, sex determination.

### Introduction

Studies on diploid male production in the haplodiploid Hymenoptera suggest that complementary sex determination (CSD) is the ancestral condition of the taxon (Cook, 1993; Cook & Crozier, 1995; Butcher et al., 2000a,b). In single locus (sl) CSD, sex is determined by the complementary action of codominant alleles at a sex-determining autosomal locus (Crozier, 1971; Bull, 1981). Under sl-CSD, heterozygotes at the sex locus develop into diploid females, hemizygotes develop into haploid males, and homozygotes develop into diploid males (Whiting, 1939). Because diploid males are effectively sterile (Stouthamer et al., 1992; Agoze et al., 1994), their production increases the genetic load, and reduces the reproductive success of their parents (Page, 1980; Ratnieks, 1990; Ross et al., 1993). In social Hymenoptera, diploid male production has been shown to increase colony mortality (Ross & Fletcher, 1986), and decrease colony growth rates (Plowright & Pallett, 1979).

There is a finite number of sex alleles in any given population, and therefore a small number of diploid males is expected to be produced by the chance mating of parents sharing a sex allele in common. Such matings

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are referred to as matched matings (Adams et al., 1977). Assuming females mate with only one male, matched matings will result in 50% of the total diploid offspring being diploid males. In a sl-CSD system, the frequency of matched matings,  $\theta$ , is expected to arise with a frequency of 2/K, where K is the effective number of alleles at the sex-determining locus (Adams et al., 1977). Thus, allelic diversity plays a critical role in controlling diploid male production. At equilibrium, K is in mutation-drift balance and increases with population size (Yokoyama & Nei, 1979). Other factors being equal, smaller populations are expected to have a lower diversity of sex alleles and, consequently, higher levels of diploid male production than larger populations. Inbreeding will also increase the frequency of matched matings and, hence, the production of diploid males in a population. This phenomenon is well documented through laboratory inbreeding, which results in the production of diploid males in hymenopteran species exhibiting CSD (e.g. Plowright & Pallett, 1979; Agoze et al., 1994; Heimpel et al., 1999; Butcher et al., 2000a). The frequency of diploid males is thus an important parameter in hymenopteran conservation genetics, as high frequencies may indicate genetic impoverishment resulting from inbreeding, population fragmentation, loss of allelic diversity, or a combination of these factors (Packer & Owen, 2001).

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Two parameters can be used to quantify diploid male production in a population. (i) The proportion of diploids that are male,  $\Phi$ : defined as the total number of diploid males divided by the total number of diploid individuals (male and female), it is equivalent to  $(1 - s)(\theta/2)$  (Adams *et al.*, 1977), where s is the selection coefficient against the production of diploid males. (ii) The proportion of males that are diploid,  $\phi$ : defined as the number of diploid males divided by the total number of males (diploid and haploid) (Owen & Packer, 1994).  $\phi$ is dependent on the frequency of matched matings and on the ratio of fertilized to unfertilized eggs (Owen & Packer, 1994). Therefore, in colonies of social Hymenoptera,  $\phi$  is expected to achieve its highest value when queens are producing workers and not haploid sons. Estimates of  $\Phi$  and  $\phi$  in natural populations usually lie below 10% (e.g. Kukuk & May, 1990; Packer & Owen, 1990). Unusually high diploid male estimates were recorded for several euglossine bees in Panama (Roubik et al., 1996), and in introduced populations of the fire ant Solenopsis invicta in the USA (Ross et al., 1993). Loss of allelic diversity at the sex locus, owing to a founder event, is responsible for high levels of diploid male production in Solenopsis invicta in its introduced range (Ross et al., 1993). Causes for high levels of diploid male production in the euglossine bees remain uninvestigated.

Halictus poeyi (Lep.) is a primitively eusocial bee found in SE U.S.A. (Packer, 1999). Morphologically it is indistinguishable from its genetically divergent sibling species *H. ligatus* (Carman & Packer, 1996; Danforth *et al.*, 1998), which is found to the north and west of the range of *H. poeyi*. The two species are sympatric only in the Piedmont along the southeastern edge of the Appalachian mountains. The present contribution documents high proportions of diploid males in samples of *H. poeyi* from Florida and presents possible explanations for this unexpected result based upon *H. poeyi*'s social biology and population dynamics.

## Materials and methods

## Collection of samples

*Halictus poeyi* samples were collected from 12 sites in Florida, U.S.A., between 6 and 21 May 2000 (Fig. 1). All bees were collected while foraging mainly on the composite flower *Bidens pilosa* between 10.00 and 15.00 hours, a period of peak bee activity. In the field, bees were killed and stored in a liquid nitrogen container, and later transferred to a  $-80^{\circ}$ C deep freezer at York University.

# Allozyme electrophoresis and diploid male detection

Allozyme electrophoresis on horizontal starch gels was used to examine genetic variation and detect diploid



**Fig. 1** Map of *Halictus poeyi* study sites in Florida, May 2000. Sample sizes are displayed between brackets.

males in *H. poeyi* samples. Enzyme staining recipes, gel recipes, gel running conditions, and scoring procedures followed Packer & Owen (1989, 1990). Males heterozygous at any locus were considered diploid. Diploid males were detected using six out of the eight polymorphic loci surveyed (Table 1).

#### Diploid male estimation

Because bees were sampled during the early stages of the colony cycle, where they are mostly producing workers, only  $\Phi$ , the proportion of diploids that are male, can be estimated with a high degree of confidence. Single locus estimates for the proportion of diploids that are male,  $\Phi$ , were calculated using the following equation (Owen & Packer, 1994):

$$\hat{\Phi} = \frac{B}{B+A},\tag{1}$$

where *B* is the total number of heterozygous males and *A* is the total number of heterozygous females per locus per sample. The frequency of matched matings  $\theta$  and the effective number of alleles at the sex determining locus *K*, assuming sl-CSD, were estimated as follows (Adams *et al.*, 1977; Owen & Packer, 1994):

$$\hat{\theta} = \frac{2\hat{\Phi}}{1-s},\tag{2}$$

$$\hat{K} = \frac{2}{\hat{\theta}},\tag{3}$$

Because the selection coefficient against diploid male s is unknown and assumed to be zero, these will be the minimum estimates of  $\theta$  and K (Owen & Packer, 1994).

#### Effective population size

The mean effective population size,  $N_e$ , was estimated using an isolation-by-distance plot (Wright, 1943; Slatkin, 1993). This was obtained by linear regression of

the log of gene flow vs. the log of distance between site pairs in Florida. Gene flow was assessed as the number of migrants per generation  $(N_{\rm m})$  using the following equation (Wright, 1951):

$$N_{\rm m} = \frac{1}{4} \left( \frac{1}{F_{\rm ST}} - 1 \right),$$
 (4)

where  $F_{ST}$  is the population subdivision statistic. Weir & Cockerham's (1984) estimator of  $F_{ST}$  was calculated from the allozyme data using the population genetics software package GENEPOP v.3.2a (Raymond & Rousset, 1995). From the isolation-by-distance plot, the mean effective population size  $N_e$  is equivalent to  $10^b$ , where b is the y-intercept of the regression line (i.e. the effective population size is equivalent to log gene flow where log distance is zero). This is also known as the neighbourhood size (Slatkin, 1993).

#### Results

#### Diploid male estimates

Diploid males were detected in five out of the 12 sites sampled in Florida (Table 2). From a total of 27 males collected in these five sites, 13 were detected to be diploid using allozyme electrophoresis. Diploid males were detected at the loci shown in Table 2.

Single locus estimates for the proportion of diploids that are male were surprisingly high across all loci and sites (Table 2). They ranged from 9.1% (JUP site-*Alddh*), to 50% (KEY site-*Dia*, etc.), and averaged 29% over all loci and sites. The frequency of matched matings ranged from 18.2% to 100%, and averaged 58%. The number of effective alleles at the sex determining locus ranged from two to 11, and averaged five alleles across all loci and sites.

#### Effective population size

The isolation-by distance plot (Fig. 2) exhibited a typical pattern of decreasing gene flow over increasing

Table 1 Enzyme names,   abbreviations, EC numbers	Enzyme	Acronym	EC number	Buffer system		
and buffer systems used	Adenylate kinase* Aldehyde dehydrogenase* Arginine kinase* Diaphorase (NADH)* Esterase Glucose-6-phosphate dehydrogenase* Hexokinase* Phosphogluconate dehydrogenase	Ak-1 Alddh Ark Dia Est-3 G6pdh Hk 6Pgd-1	2.7.4.3 4.1.2.13 2.7.3.3 1.8.1.n 3.1.1.1 1.1.1.49 2.7.1.1 1.1.1.43	CAM V CAM V RSL CAM I CAM		

\*Indicates enzymes that detected at least one diploid male.

Site	Diploid/ total √	Locus	$\Phi$ (± SD)	θ	K
FOR	3/9	Ak-1	$0.333 \pm 0.272$	0.667	3
	,	Ark	$0.333 \pm 0.272$	0.667	3
		G6pdh	$0.500 \pm 0.354$	1	2
		Ηk	$0.500 ~\pm~ 0.354$	1	2
LOR	4/8	Alddh	$0.231 \pm 0.117$	0.462	4
	,	Ark	$0.500 \pm 0.354$	1	2
		Dia	$0.250\ \pm\ 0.217$	0.500	4
FRU	1/1	Alddh	$0.143 \pm 0.132$	0.286	7
JUP	3/6	Alddh	$0.091~\pm~0.087$	0.182	11
	,	Dia	$0.333 \pm 0.272$	0.667	3
		G6pdh	$0.143\ \pm\ 0.132$	0.286	7
KEY	2/3	Alddh	$0.125 \pm 0.117$	0.250	8
	,	Dia	$0.500 \pm 0.354$	1	2
		Hk	$0.167\ \pm\ 0.152$	0.333	6
Range (%)			9.1–50	18.2-100	2-11
Average (%	)		29	58	5

**Table 2** Single locus estimates of the proportion of diploids that are male  $(\Phi)$ , the frequency of matched matings  $(\theta)$ , and the effective number of alleles at the sex determining locus (K), for sites that contained diploid males

geographical distance. The regression has a significant y-intercept (t = 3.201209, P = 0.0034) of  $1.29 \pm 0.40$  (SE), equivalent to a mean effective population size of 19.6  $\pm$  2.5 (SE) individuals.

#### Discussion

Diploid male production is a by-product of CSD. Because diploid males are effectively sterile, their production is disadvantageous (Page, 1980; Agoze *et al.*, 1994; Ratnieks, 1990 Stouthamer *et al.*, 1992), and thus they are not expected to occur at a high frequency in natural populations. Most studies coroborate this expectation (Kukuk & May, 1990; Packer & Owen, 1990; reviewed by Cook, 1993). Nonetheless, several researchers have found diploid males at high frequencies. Roubik *et al.* (1996) found that the proportion of males that



**Fig. 2** Linear regression between the logarithm of migrants per generation (Nm) and the logarithm of distance (D) for *Halictus poevi* in Florida, 2000.

were diploid,  $\phi$ , ranged from 12 to 100% in their samples of euglossine bees in Panama. The causes for such high levels of diploid male production awaits further empirical investigations. Unfortunately, as females of these orchid bees are not attracted to chemical baits as the males are, it was not possible for Roubik et al. to calculate the proportion of diploids that are male, the statistic required in order to estimate the frequency of matched matings and the effective number of sex determining alleles. Ross et al. (1993) also discovered unusually high levels of diploid male production in introduced populations of the fire ant Solenopsis invicta in the U.S.A., with  $\phi$  ranging from 73.3 to 100%. This was attributed to the loss of allelic diversity at the sexdetermining locus during a recent founder event that led to the invasion of the fire ant in the U.S.A.

High proportions of diploid males were found in populations of *Halictus poeyi* from Florida, where it is a common and widespread species. This combination of high levels of diploid male production in an apparently abundant bee is somewhat enigmatic and requires explanation. There are four likely factors that contribute to this paradox.

1 Like many ground-nesting bees, *H. poeyi* prefers sparsely vegetated nonsandy soil, and such early successional stage habitats are uncommon and temporary, particularly in warm, damp Florida. It is well known that frequent extinction and recolonization events have a highly negative influence on levels of genetic variability (Giplin, 1991), and can reduce the effective population size by several orders of magnitude (Maruyama & Kimura, 1980; McCauley, 1993).

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2 Florida has been influenced by repeated sea-level fluctuations, which have, at times, flooded much of the state, causing repeated fragmentation and isolation of animal and plant populations (e.g. Ellsworth *et al.*, 1994; references therein). When combined with the extinction/recolonization population ecology of this species, this would further reduce the effective population size and opportunity for maintenance of large numbers of sex determining alleles.

**3** The large-scale habitat destruction resulting from extensive agriculture and the associated use of pesticides will also have resulted in the reduction and fragmentation of populations since European settlement over 200 hundred years ago.

These three factors all suggest that H. poeyi has a population structure in Florida which is likely to result in low allelic diversity at the sex locus. One other aspect of its biology suggests that its effective population size may not be as high as it might appear in the field.

**4** As a social bee that can build up reasonably large colony sizes from solitarily initiated nests (Packer & Knerer, 1986), *H. poeyi* will often appear far more abundant than its effective population size would suggest as a result of the large number of workers per nest.

Taken together, these factors suggest that the estimated effective population size obtained herein for this species – in the order of around 20 individuals – is not necessarily surprising even though the species can be quite abundant. What is more, because of the aforementioned reasons, such small effective populations may have been in existence for a long period of time.

The calculated average of only five alleles at the sex locus in *H. poeyi* is not so surprising in light of the factors noted above. Although the number of alleles at the sex locus in natural hymenopteran populations usually ranges between nine and 20 (reviewed by Cook & Crozier, 1995), two other studies have found similarly low diversity. Heimpel *et al.* (1999) estimated there to be six sex alleles in a very small population of the parasitoid wasp *Bracon hebetor*. Similarly Butcher *et al.* (2000a,b) found three to four sex alleles for the parasitic wasps *Diadegma fabricianae* and *D. armillata* in small and isolated populations near the northern edge of their natural range.

The existence of diploid males in nature poses an additional problem to the conservation genetics of haplodiploid insects such as the parasitoid wasps and the bees (Packer & Owen, 2001), two groups of considerable importance for biological control and pollination, respectively. Yet there has been little attention paid to this subject. The high proportion of males found to be diploid in a common, primitively eusocial bee suggests a simple means of gathering more data on this

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topic. Primitively eusocial species with an annual colony cycle, such as almost all social Halictidae, bumblebees and yellowjackets, produce diploids for much of the colony cycle, switching to male and reproductive female production late in the summer (e.g. Packer & Knerer, 1987). Thus, it is predicted that many of the males found during the early summer in these species should be diploid, and if their frequency as a proportion of all diploids can be estimated without bias (e.g. through censuses of brood in nests), accurate measurements of the allelic diversity at the sex locus could be obtained.

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## References

- ADAMS, J., ROTHMAN, E. D., KERR, W. E. AND PAULINO, Z. L. 1977. Estimation of the number of sex alleles and queen matings from diploid male frequencies in a population of *Apis mellifera*. *Genetics*, **86**, 583–596.
- AGOZE, M. E., DREZEN, J. M., RENAULT, S. AND PERIQUET, G. 1994. Analysis of the reproductive potential of diploid males in the wasp *Diadromus pulchellus* (Hymenoptera: Ichneumonidae). *Bull. Ent. Res.*, **84**, 213–218.
- BULL., J. J. 1981. Coevolution of haplo-diploidy and sex determination in the Hymenoptera. *Evolution*, 35, 568–580.
- BUTCHER, R. D. J., WHITFIELD, W. G. F. AND HUBBARD, S. F. 2000a. Single-locus complimentary sex determination in *Diadegma chrysostictos* (Gmelin) (Hymenoptera: Ichneumonidae). *J. Hered.*, **91**, 104–111.
- BUTCHER, R. D. J., WHITFIELD, W. G. F. AND HUBBARD, S. F. 2000b. Complementary sex determination in the genus *Diadegma* (Hymenoptera: Ichneumonidae). J. Evol. Biol., **13**, 593–606.
- CARMAN, G. M. AND PACKER, L. 1996. A cryptic species allied to *Halictus ligatus* Say (Hymenoptera: Halictidae) detected by allozyme electrophoresis. *J. Kans. Entomol. Soc.*, **69**, 168–176.
- COOK, J. M. 1993. Sex determination in the Hymenoptera: a review of models and evidence. *Heredity*, **71**, 421–435.
- COOK, J. M. AND CROZIER, R. H. 1995. Sex determination and population biology in the Hymenoptera. *Trends Ecol. Evol.*, **10**, 281–286.
- CROZIER, R. H. 1971. Heterozygosity and sex determination in haplodiploidy. Am. Nat., 105, 399–412.
- DANFORTH, B. N., MITCHELL, P. L. AND PACKER, L. 1998. Mitochondrial DNA differentiation between two cryptic *Halictus* (Hymenoptera: Halictidae) species. *Ann. Entomol. Soc. Am.*, **91**, 388–391.

- ELLSWORTH, D. L., HONEYCUTT, R. L., SILVY, N. J., BICKHAM, J. W. AND KLIMSTRA, W. D. 1994. Historical biogeography and contemporary patterns of mitochondrial DNA variation in white-tailed deer from the southeastern United States. *Evolution*, **48**, 122–136.
- GIPLIN, M. 1991. The genetic effective size of a metapopulation. Biol. J. Linn. Soc., 42, 165–175.
- HEIMPEL, G. E., ANTOLIN, M. F. AND STRAND, M. R. 1999. Diversity of sex-determining alleles in *Bracon hebetor*. *Heredity*, **82**, 282–291.
- KUKUK, P. F. AND MAY, B. 1990. Diploid males in a primitively eusocial bee, *Lasioglossum (Dialictus) zephyrum* (Hymenoptera: Halictidae). *Evolution*, **44**, 1522–1528.
- MARUYAMA, T. AND KIMURA, M. 1980. Genetic variation and effective population size when local extinction and recolonization of subpopulations are frequent. *Proc. Natl. Acad. Sci. U.S.A.*, **77**, 6710–6714.
- MCCAULEY, D. E. 1993. Genetic consequences of extinction and recolonization in fragmented habitats. In: Kareiva, P. M., Kingsolver, J. G. and Huey, R. B. (eds) *Biotic Interactions* and Global Change, pp. 217–233. Sinauer, Sunderland, MA.
- OWEN, R. E. AND PACKER, L. 1994. Estimation of the proportion of diploid males in populations of Hymenoptera. *Heredity*, **72**, 219–227.
- PACKER, L. 1999. The Distribution of *Halictus Ligatus* Say and *H. Poeyi* Lep. (Hymenoptera: Halictidae) in North America. *Spec. Publ. Univ. Kansas Nat. Hist. Mus.*, **24**, 81–84.
- PACKER, L. AND KNERER, G. 1986. The biology of a subtropical population of *Halictus ligatus* Say (Hymenoptera; Halictidae) I. Phenology and social organization. *Behav. Ecol. Sociobiol.*, 18, 363–375.
- PACKER, L. AND KNERER, G. 1987. The biology of a subtropical population of *Halictus ligatus* Say (Hymenoptera; Halictidae) III. The transition between annual and continuously brooded colony cycles. J. Kans. Entomol. Soc., 60, 510–516.
- PACKER, L. AND OWEN, R. E. 1989. Allozyme variation in *Halictus Rubicundus* (Christ): a primitively social halictine bee (Hymenoptera: Halictidae). *Can. Ent.*, **121**, 1049–1058.
- PACKER, L. AND OWEN, R. E. 1990. Allozyme variation, linkage disequilibrium and diploid male production in a primitively social bee *Augochlorella striata* (Hymenoptera; Halictidae). *Heredity*, **65**, 241–248.

- PACKER, L. AND OWEN, R. E. 2001. Population genetic aspects of pollinator decline. *Conserv. Ecol.*, 5: 4 (www.consecol.org/ vol5/iss1/art4,
- PAGE, R. E. 1980. The evolution of multiple mating behavior by honey bee queens (*Apis mellifera*). *Genetics*, **96**, 263–273.
- PLOWRIGHT, R. C. AND PALLETT, M. J. 1979. Worker-male conflict and inbreeding in bumble bees (Hymenoptera: Apidae). *Can. Ent.*, **111**, 289–294.
- RATNIEKS, F. L. W. 1990. The evolution of polyandry by queens in social Hymenoptera: the significance of the timing of removal of diploid males. *Behav. Ecol. Sociobiol.*, **26**, 343–348.
- RAYMOND, M. AND ROUSSET, F. 1995. GENEPOP (Version 1.2): population genetics software for exact tests and ecumenicism. J. Hered., 86, 248–249.
- ROSS, K. G. AND FLETCHER, D. J. C. 1986. Diploid male production – a significant colony mortality factor in the fire ant *Solenopsis invicta* (Hymenoptera: Formicidae). *Behav. Ecol. Sociobiol.*, **19**, 283–291.
- ROSS, K. G., VARGO, E. L., KELLER, L. AND TRAGER, J. C. 1993. Effect of founder even on variation in the genetic sexdetermining system of the fire ant *Salenopsis invicta*. *Genetics*, 135, 843–854.
- ROUBIK, D. W., WEIGT, L. A. AND BONILLA, M. A. 1996. Population genetics, diploid males, and limits to social evolution of eugolossine bees. *Evolution*, **50**, 931–935.
- SLATKIN, M. 1993. Isolation by distance in equilibrium and non-equilibrium populations. *Evolution*, **47**, 264–279.
- STOUTHAMER, R., LUCK, R. F. AND WERREN, J. H. 1992. Genetics of sex determination and the improvement of biological control using parasitoids. *Environ. Entomol.*, 21, 427–435.
- WEIR, B.S. AND COCKERHAM, C.C. 1984. Estimating F-statistics for the analysis of population structure. *Evolution*, **38**, 1358–1370.
- WHITING, P. W. 1939. Sex determination and reproductive economy in *Habrobracon. Genetics*, 24, 110–111.
- WRIGHT, s. 1943. Isolation by distance under diverse systems of mating. *Genetics*, **31**, 39–59.
- WRIGHT, s. 1951. The genetical structure of populations. Ann. Eugen., 15, 323–354.
- YOKOYAMA, S. AND NEI, M. 1979. Population dynamics of sexdetermining alleles in honey bees and self-incompatibility alleles in plants. *Genetics*, **91**, 609–626.